



## COMPARATIVE EVALUATION OF *IN VITRO* ANTIBACTERIAL AND ANTIOXIDANT ACTIVITY USING STANDARD DRUG AND POLYHERBAL FORMULATION

S. Vetrivelvan<sup>1†</sup>, J. Shankar<sup>2\*</sup>, S. Gayathiri<sup>2</sup>, S. Ishwin<sup>2</sup>, C. Hemah Devi<sup>2</sup>, A. Yaashini<sup>2</sup>, G. Sheerenjet<sup>2</sup>

<sup>1</sup>Masterskill University College of Health Sciences, Cheras, Selangor, Malaysia.

<sup>2</sup>Faculty of Science Technology and Engineering, La Trobe University, Bendigo, Australia.

### ABSTRACT

The present study was taken to investigate the *in vitro* antibacterial and antioxidant activity of polyherbal formulation comparative to standard drug and individual plant extract. Cup and Plate method was used for *in vitro* antibacterial screening. Zones of inhibition were observed in disc diffusion for antibacterial investigation against Gram-positive and Gram negative pathogenic bacteria. The Polyherbal formulation showed average zone of inhibition comparative to individual plant extract and standard drug ranged from 9-12 mm. In antioxidant screening, the compound showed significant.

**Keywords:** Polyherbal formulation, Cup and Plate method, Antibacterial and Antioxidant activity, FRAP assay.

### INTRODUCTION

Medicinal plants are widely exploited worldwide for their active ingredients. This has created a wide gap between the production and exploitation of most of the medicinal plants of important plant species.

The Siddha System of Medicine (Traditional Tamil System of medicine), which has been prevalent in the ancient Tamil land, is the foremost of all other medical systems in the world. The uniqueness of Siddha System is evident by its continuous service to the humanity for more than 5000 years in combating diseases and also in maintaining its physical, mental and moral health while many of its contemporaries had become extinct long ago. Aromatic and medicinal plants are known to produce certain bioactive molecules which react with other organisms in the environment, inhibiting

bacterial or fungal growth. The substances that can inhibit pathogens and have little toxicity to host cells are considered candidates for developing new antimicrobial drugs. Spices and herbs have been used for thousands of centuries by many cultures to enhance the flavor and aroma of foods. Scientific experiments since the late 19<sup>th</sup> century have documented the antioxidant properties of some spices, herbs, and their components (Zaika, 1988; Bajpai *et al.*, 2005). Many studies reported the activities of spices and herbs on food borne pathogenic microorganisms (Arora and Kaur, 1999; Yano *et al.*, 2006). As such developing a polyherbal formulation with definitely produce synergistic effect as needed comparable to standard drugs that are available in market all over the world. The purpose of the present study was to investigate the antioxidant and antimicrobial properties of *Withania somnifera*, *Bacopa monnieri* and *cinnamomum zeylanicum* individually and combination of all this three as polyherbal treatment. In this paper we report the results of such studies in order to orient future investigations towards the finding of new, potent and safe antioxidant and antimicrobial compounds.

Corresponding Author

S. Vetrivelvan

Email:- [vetricology@gmail.com](mailto:vetricology@gmail.com)

*Withania somnifera* is a hedge plant that blooms in India and also the North America. The roots of the ashwagandha plant have been in use for treatment by Ayurvedic healers. It contains phyto-androgens, which are the precursors to reproductive hormones. Furthermore, this plant helps in reproductive health for men and women. The active constituents are slightly sedative in nature and it calms the central nervous system. It is reported that ashwagandha shows a great effect in reducing inflammation, increasing mental activity, invigorating the body, decreasing stress and as an antioxidant.

*Bacopa monnieri* is the small herb with the numerous branches. It grows to a height of 2 -3 feet and its branches are 10 -35 cm long. It has oval shaped leaves that are 1-2 cm long and 3- 8 mm broad. Leaves are formed in pairs along the stems. Small- tubular, five pedaled flowers are white- purple in colour. Its stem is soft, succulent, and hairy with the glands. Roots emerge out of the nodules and directly go to the soil. The fruit is oval and sharp at apex. It contains triterpene, saponins of the dammarane class, which are bacosides and bacopasaponins, and which contain 2 or 3 sugars each. Other constituents include mannitol, common plant sterols, and betulinic acid, as well as glutamic and aspartic acids. It is reported that brahmi shows a great effect in treating asthma, hoarseness, insanity, epilepsy, nerve tonic, cardio tonic and also diuretic.

*Cinnamon* common name is true cinnamon and its scientific name is *cinnamomum zeylanicum*. The ornamental is of a tree with golden red bark that is dried and is the cinnamon spice. It can grow well in soil which contains lot of sand and it grows at tropical climate. New foliage is deep red, and small white flowers are followed by dark purple fruit. The leaves contain eugenol and are sometimes used as a substitute for cloves and represent the major aromatic chemical component of cloves. Native region is from Sri Lanka, and Burma. The useful chemical component is cinnamon's bark is its oil which is just 4 % but others are cinnamaldehyde (3-phenyl-acrolein, 65 to 75%) and eugenol (4-(1-propene-3-yl)-2-methoxyphenol, 5 to 10%) which is also determine the flavor and aroma. It is proven to have many pharmacological effects such as anti microbial, anti-inflammatory, antioxidant, and also used for type 2 diabetes.

## MATERIALS AND METHODS

### Preparation of Formulation

Readymade extracts of *Withania somnifera*, *Bacopa monnieri* and *cinnamomum zeylanicum* extract is obtained from department of ayurveda, Banarus Hindu University, Varanasi. Use to determine their antibacterial and antioxidant activity.

Comparison between individual plant extract, poly herbal formulation and standard drug has been done using cup and plate method. The standard pathogenic bacteria cultures were used in this study. The bacterial cultures were rejuvenated in Mueller- Hinton broth at 37 degree Celsius for 24 hours. The pathogenic bacterial culture was inoculated into sterile Nutrient broth and incubated at 37 degree Celsius for 3 hours.

### Preparation of poly herbal formulation

Polyherbal formulation was prepared by simple percolation process. Distilled water was used to make up the volume up to 5ml.

All of the chemicals used in this work were obtained from Masterskill University College of Health Sciences Laboratory. The chemicals were analytical degree.

### Biological materials

Three bacteria cultures were used in the study: *Klebsiella aerogenes*, *Pseudomonas aeruginosa* and *Escherichia coli*. The microorganisms maintained on Nutrient Agar and Muller Hinton (Merck, Darmstadt, Germany) were supplied by Microbiology Laboratory of Masterskill University College of Health Sciences.

### Antibacterial Screening

#### Antibacterial Activity using cup and plate method

The cup and plate method method was employed to determine the antibacterial activity of poly herbal preparations. Turbidity of inoculums was matched with McFarland turbidity standard. Inoculums were spread over the Nutrient agar plate using a sterile cotton swab in order to get a uniform microbial growth. Then the prepared antibacterial disc were placed over the lawn and pressed slightly along with positive and negative controls. Vancomycin antibiotic disc were used. The plates were incubated for 24 hours at 37 °C. The antibacterial activity was evaluated for 0.2 ml of each plant extract and polyherbal extract. The diameter of inhibition zones were measured and recorded.

### In vitro antioxidant activity

#### Ferric Ion Reducing/antioxidant Power Assay (FRAP)

Antioxidant activity was determined by Ferric ion reducing antioxidant power assay (FRAP) as described by Oyaizu. 2ml of different plant extract (40mg/ml) were mixed with phosphate buffer (2.5 ml, 0.2 M, pH 6.6) and 0.1 % potassium ferricyanide (2.5 ml). The mixture was incubated at 50 °C for 20 min. Aliquots of 10 % trichloroacetic acid (2.5 ml) were added to the mixture, which was then centrifuged at 3000 rpm for 10 min. The upper layer of solution (2.5 ml) was mixed with distilled water (2.5 ml) and a freshly prepared ferric

chloride solution (0.5 ml, 0.1%). The absorbance was measured at 640 by using UV Spectrophotometer.

#### Statistical evaluation

The antimicrobial activity was determined by measuring the diameter of zone of inhibition that is the mean of two replicates.

### RESULTS AND DISCUSSION

**Table 1. Composition of Polyherbal Formulation**

Botanical name	Weight	Parts used
<i>Withania somnifera</i>	100mg	<b>whole plant</b>
<i>Bacopa monnieri</i>	100mg	<b>whole plant</b>
<i>Cinnamomum zeylanicum</i>	100mg	<b>whole plant</b>

**Table 2. In vitro antibacterial activity of Individual plant extract, polyherbal formulation and standard drug**

Test organism	Diameter of zone of inhibition (in mm)				
	BE	WS	CZ	PH	VN
<b>Gram positive bacteria</b>					
<i>Klebsiella aerogenes</i>	<b>16</b>	<b>17</b>	<b>17</b>	<b>19</b>	<b>23</b>
<b>Gram negative bacteria</b>					
<i>Pseudomonas aeruginosa</i>	<b>16</b>	<b>14</b>	<b>16</b>	<b>17</b>	<b>22</b>
<i>Escherichia coli</i>	<b>17</b>	<b>17</b>	<b>16</b>	<b>16</b>	<b>24</b>

**Table 3. Antioxidant activity of polyherbal formulation**

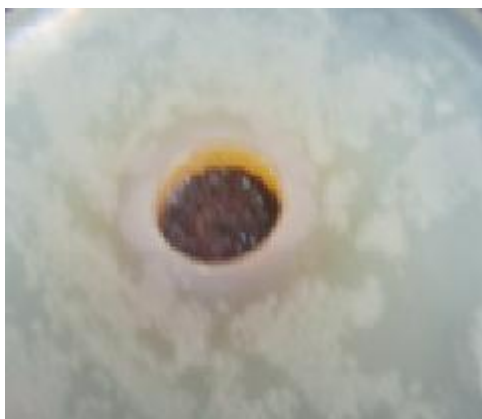
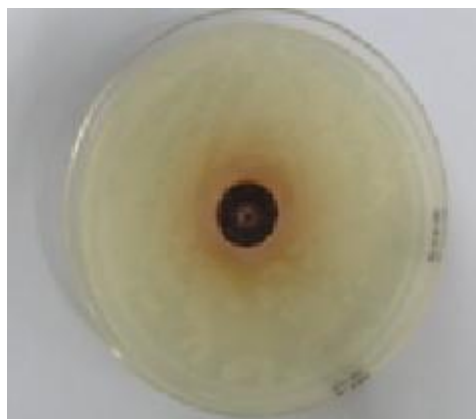
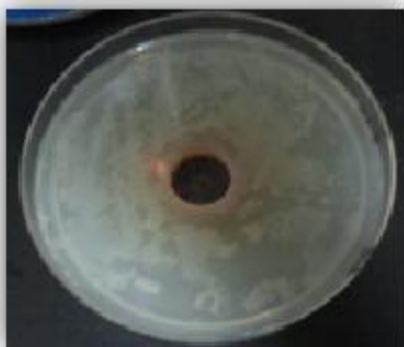
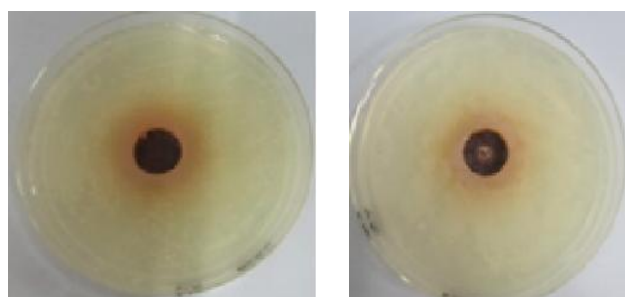
Concentration in µg/ml	Absorbance at 640 nm
100	0.63
200	0.68
300	0.93
400	1.07
500	1.10

**Figure 1. Standard antibiotic disc before incubation**



**Figure 2. Zone of inhibition of standard antibiotics**



**Figure 3. Zone of inhibition of Brahmi Extract****Figure 4. Zone of inhibition of Cinnamon Extract****Figure 5. Zone of inhibition of Aswagandha Extract****Figure 6. Greater inhibition zone of polyherbal formulation in Pseudomonas aeruginosa and Klebsiella aerogenes**

## CONCLUSION

The present study showed that Polyherbal formulation has got antibacterial and antioxidant activity. The Polyherbal formulation showed moderate to mild antibacterial activity against most of the tested bacteria. It may be concluded that Polyherbal formulation is active against the tested microorganisms and have good antioxidant activity. Further studies should be done to study on toxicity and clinical studies in order to develop a future drug containing this poly herbal formulation for treatment of infection and cancer.

## ACKNOWLEDGEMENT

The authors are grateful to the Faculty of Science Technology and Engineering, La Trobe University, Bendigo, Australia and Faculty of Pharmacy, Masterskill University College of Health Sciences, Malaysia for supporting and providing access to free online journals to conduct this research. The authors are also wish to thank MUCH Laboratory assistant's in providing materials and helping in conducting this work.

## Declaration of interest

The authors report no conflicts of interest.

## REFERENCES

- Anthony JM, Senaratna T, Dixon KW, Sivasithamparam K. The Role of Antioxidants for initiation of Somatic Embryos with *C. onostephim pendulum* (Ericaceae) plant cell tissue. *Org. Cult*, 78, 2004, 247-252.
- Bouamama H, Noel T, Villard J, et al. Antimicrobial activities of the leaf extracts of two Moroccan *Cistus L.* species. *Journal of Ethnopharmacology*, 104, 2006, 104-107.
- Darokar MP, Khanuja SPS, Shasany AK, Kumar S. Low levels of genetic diversity detected by RAPD analysis in geographically distinct accessions of *Bacopa monnieri*. *Genetic Resources and Crop Evolution*, 48, 2001, 555-558.
- Habsah M, Amran M, Mackeen MM, Lajis NH, Kikuzaki H, Nakatani N, et al. Screening of Zingiberaceae extracts for antimicrobial and antioxidant activities. *Journal of Ethnopharmacol*, 72, 2000, 403-10.

- Ibrahim H, Khalid N, Hussin K. Cultivated gingers of Peninsular Malaysia: utilization, profiles and micropropagation. *Gard Bull Sing*, 59, 2007, 77-88.
- Kikuzaki H, Nakatani N. Antioxidant effects of some ginger constituents. *Journal of Food Science*, 58(6), 1993, 1407-1410.
- National Committee for Clinical Laboratory Standards, Performance Standards for antimicrobial susceptibility testing, 8th Informational Supplement, Villanova, Pa, M100S12, 2002.
- Okemo PO, Bais HP, Vivanco JM. In vitro activities of *Maesa lanceolata* extracts against fungal plant pathogens. *Fitoterapia*, 74, 2003, 312-316.
- Oyaizu M. Studies on product browning reaction prepared from glucosamine. *J. Nutri*, 44, 1986, 307-315.
- Pietta PG. Flavonoids as antioxidant. *J. Nat. Prod*, 63, 2000, 1035-1042.
- Tushar, Basak S, Sarma GC, Rangan L. Ethnomedical uses of Zingiberaceous plants of Northeast India. *J Ethnopharmacol*, 132, 2010, 286-96.
- Vasala PA. Ginger. In: Peter KV, Editor. *Handbook of Herbs and Spices*. Woodhead Publishing Ltd. and CRC Press. 2001. p. 195-206.
- Yen GC and Duh PD. Scavenging effect of methanolic extracts of peanut hulls on free radical and active-oxygen species. *J Agric Food Chem*, 42, 1994, 629-632.
- Zampini IC, Cuello S, Alberto MR, et al. Antimicrobial activity of selected plant species from the Argentine puna against sensitive and multiresistant bacteria. *Journal of Ethnopharmacology*, 124, 2009, 499-505.